

Instructions for using PrimeSurface® 96 Slit-well Plate

Notes

- This instruction explains the basic operating procedure for PrimeSurface® 96 Slit-well Plate.
- Instructions recommended here are the best operating protocols but depending upon your cell line and assay, some optimizations may be required.
- The product and instruction are subject to change.
- This product is for research purpose only.

Features

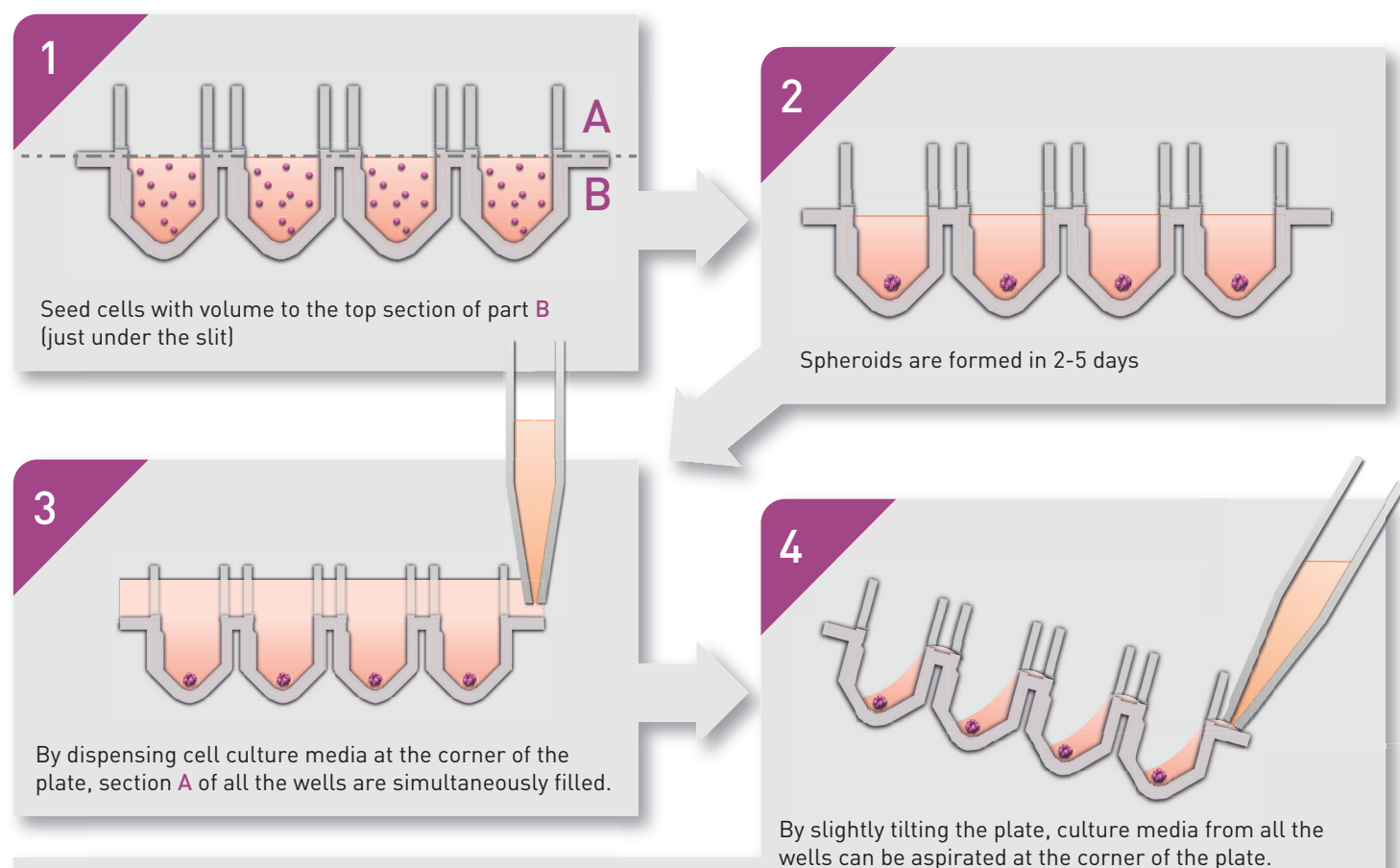
- PrimeSurface® 96 Slit-well Plate, is designed to form and maintain uniform spheroid for long term cultures.
- New and innovative design of plate with slits on the upper half of each well.

- Media aspirating and dispensing is efficiently performed from the corners of the plate while keeping the spheroid formation undisturbed in wells.

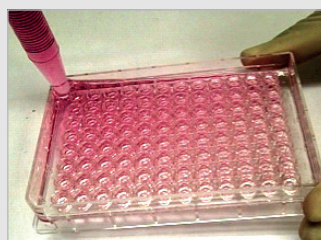
Description

- The figures below provide a schematic explanation for the use of PrimeSurface® 96 Slit-well Plate in spheroid formation and media exchange.
- Procedures of cell spheroid formation (Figs. 1-3) is described in detail on page 2.
- Procedure for exchanging culture media (Fig. 4) is described on page 2.

Spheroid formation procedure using PrimeSurface® 96 Slit-well Plate



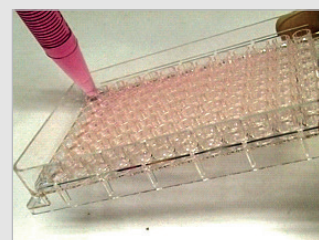
Place the pipette tip at one corner of the plate



Start aspirating the culture media slowly



Continue to tilt the plate gradually while aspirating the media



Aspirating media in this manner keeps the spheroids in the wells

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Procedures of cell spheroid formation

1. Open the packaging of a new Slit-well Plate under the hood.
2. Seed cells in the plate at a density of 6,000-9,000 cells/100 μ L/well.
 - The recommended volume of the above cell suspension is approximately 100 μ L.
 - As the maximum capacity of each well is 100 μ L, do not exceed 100 μ L volume for cell suspension per well.
 - The formed spheroids are apt to slip through the slit or protrude outside the well if the number of seeded cells is insufficient.
 - Dispense the cell suspension at the bottom of each well (section B). (Do not dispense it in section a of the well, i.e. in the slit section).
3. Culture the cells in the incubator.
 - After every 24-48 hrs, check for spheroid formation.
4. Dispense fresh culture media (20 mL per plate) at the corner of the plate using a 25 mL pipette.
 - The total volume of the dispensed culture media is 30 mL per plate, i.e. an aliquot of 10 mL during cell seeding, plus 20 mL in this step (as stated above).
 - Maximum capacity of the plate is 40 mL/plate.
 - Dispense media slowly (it should take about 20 seconds to dispense 20 mL volume to the plate).
5. Gently shake the plate back and forth in all directions a couple of times to ensure that all the wells have been evenly covered with media.
6. Place the plate in the incubator.

Procedure of exchanging culture media

1. Take the plate out of the incubator and check the spheroid formation under a microscope before exchanging the culture media.
2. Aspirate the media from a corner of the plate with a 25 mL pipette and dispose of it.
 - Apply the pipette tip to the corner of the plate and gently aspirate the old culture media while slightly tilting the plate.
 - The plate must be tilted in order to properly aspirate the culture media.
 - The recommended aspirating volume is 15 to 20 mL (takes about 20 seconds).
3. Dispense fresh media from the corner of the plate with a 25 mL pipette. This volume should be the same as that of aspiration step (2).
 - Note that the capacity of the slit plate is 40 mL/plate. It is advised to stay with total volume of 30 mL/plate.
 - Dispense media slowly (takes 20 sec) to avoid any overflow.
4. Gently shake the plate back and forth in all directions a couple of times to ensure that all the wells and corners of the plate have been evenly covered with media.

Supporting Information

For further directions on exchanging culture medium, refer to the video.

<https://www.phchd.com/eu/biomedical/cell-culture-plates/primesurface-96-slit-well>



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