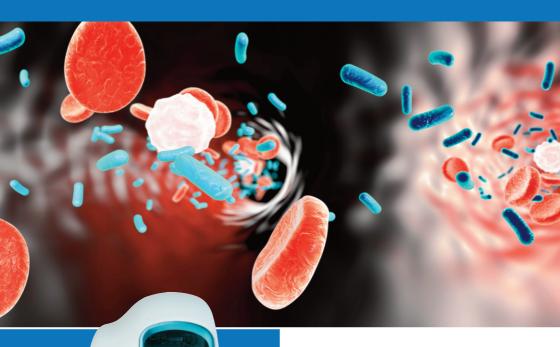


PATHFAST™ PRESEPSIN



LIFE SAVING
Presepsin The Sepsis Biomarker

A short monograph

EARLY DIAGNOSIS & PROGNOSIS

PATHFAST



Contents

Sepsis is a worldwide healthcare problem	
What is sepsis?	
Sepsis diagnosis: Time is survival	
Medical scores and biomarkers for sepsis	
What is Presepsin?	1
Measurement of Presepsin	1
How to interpret Presepsin values?	1
Specifity and sensitivity of Presepsin	1
Early sepsis marker	1
Low influence from burns, trauma and medical surgery	1
Prognosis and monitoring with Presepsin	2
Organ dysfunction and Presepsin	2
Pathogens and Presepsin	2
Presepsin and invasive fungal infections	2
Application of Presepsin in a clinical setting	3
Neutropenia	3
Perioperative risk assessment	3
Pneumonia	3
Other infectious diseases including local infections	3
Other diseases that might affect Presepsin levels	3
Neonatals	4
Conclusions	4

Sepsis is a worldwide healthcare problem

Sepsis is estimated to affect more than 30 million people worldwide every year, potentially leading to 6 million deaths (1). The burden of sepsis is most likely highest in low- and middle-income countries. Sepsis is the most common cause for readmissions to the hospital costing more than \$2 billion per year (2). The number of hospital admissions for sepsis following healthcare-associated and community-acquired infections increased up to three-fold over the last decade. In comparison, hospital admissions for stroke and myocardial infarction remained stable over the same period (**Fig. 1**; 3). Sepsis is the number 1 cost of hospitalization in the U.S. consuming

more than \$24 billion each year. The average cost per hospital stay for sepsis is \$18,400, double the average cost per stay compared to all other conditions (4) Moreover, it is estimated that 3 million newborns and 1.2 million children suffer from sepsis globally every year (5). Three out of ten deaths due to neonatal sepsis are thought to be caused by resistant pathogens (6). Furthermore, one in ten deaths associated with pregnancy and childbirth is due to maternal sepsis with over 95% of deaths occurring in low- and middle-income countries (7). Totally, one million newborn deaths are associated with maternal infection, such as maternal sepsis, each year (8).

What is sepsis?

Sepsis is a serious medical condition caused by an overwhelming immune response to infection. The body releases immune chemicals into the blood to combat the infection. Those substances trigger widespread inflammation, which leads to blood clots and leaky blood vessels. As a result, blood flow is impaired, organs are deprived of nutrients and oxygen and organ damage can be the final outcome. Common symptoms of sepsis are fever, chills, rapid breathing and heart rate, rash, confusion, and disorientation (9).

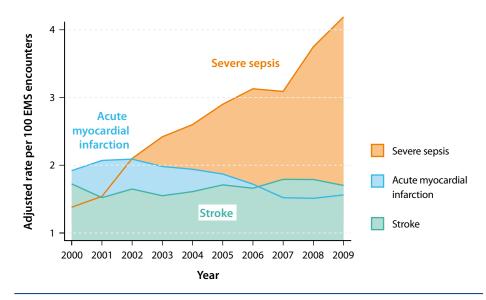
In 1991, an earlier sepsis definition, Sepsis-1, was established as "systemic inflammatory response to documented/suspected infection. Patients who met two or more of the SIRS criteria fulfilled the definition of the SIRS. Sepsis which is complicated by organ dysfunction was termed "severe sepsis", which could progress to "septic shock" (10).

A 2001 SSC task force expanded the list of diagnostic criteria, resulting in the introduction of

the Sepsis-2 definition (4). However, the definitions of sepsis and septic shock remained unchanged for more than two decades (11).

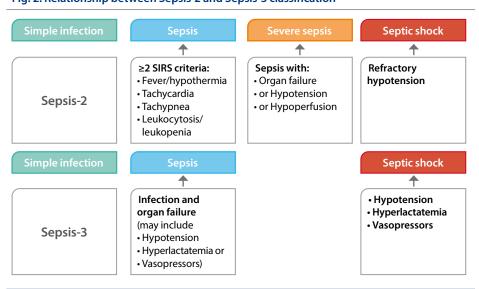
In 2016 the task force compared traditional SIRS criteria to other methods, including the Logistic Organ Dysfunction System (LODS) and Sequential Organ Failure Assessment (SOFA) scoring. The authors recommended the use of SOFA scoring to assess the severity of organ dysfunction in a potentially septic patient and redefined sepsis, known as "Sepsis-3" and the SOFA score used as criteria for the diagnosis of sepsis. Since 2016, SCCM/ESICM recommended a simplified method termed "quick SOFA" to facilitate fast identification and risk assessment of patients at admission. To build up a SOFA score takes time and needs additional laboratory data and is therefore not easy to use for urgent clinical decisions. The definition and diagnosis criteria for "septic shock" were also revised (Fig. 2; 12, 13).

Fig. 1: Hospital admissons for sepsis in comparison to Stroke or Acute myocardial infarction (AMI)



Adapted from Seymor et al., 2012 (3)

Fig. 2: Relationship between Sepsis-2 and Sepsis-3 classification



SIRS: systemic inflammatory response syndrome. Adapted from Carneiro et al., 2017 (13)

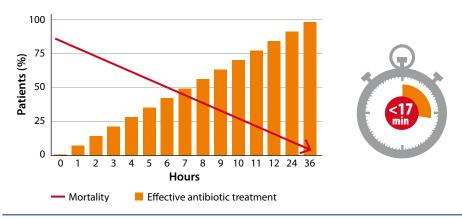
Sepsis diagnosis: Time is survival

Management of sepsis is a complicated clinical challenge requiring early recognition and management of infection, hemodynamic complications and other organ dysfunctions. The Surviving Sepsis Campaign (SSC) is a joint international program to reduce mortality caused by sepsis (11). Delays in sepsis recognition and slow initiation of treatment in multiple settings have been associated with worse outcomes, while early evidence-based treatment has been shown to improve survival (Fig. 3; 3, 12, 14). As the Surviving Sepsis Campaign's messages evolved, the 1-hour bundle of care treatment has been introduced being a valuable tool for caregivers' application upon recognition of sepsis/septic shock using a new diagnostic criteria (15). This bundle was reduced from 6 and 3 hours to finally 1 hour to enable more rapid interventions for adult sepsis and septic shock patients. Initiation of the sepsis treatment is critical to reduce mortality from sepsis and septic shock (Fig 4; 15).

The 1-hour bundle consists of the measurement of lactate and the obtaining of blood cultures prior to the administration of antibiotics, broad-spectrum antibiotics, or the application of intravenous fluids and vasopressors. A rapid and reliable biomarker at the point of care for the detection of sepsis which is applicable directly in the ICU or ER can support the diagnosis of sepsis (15).

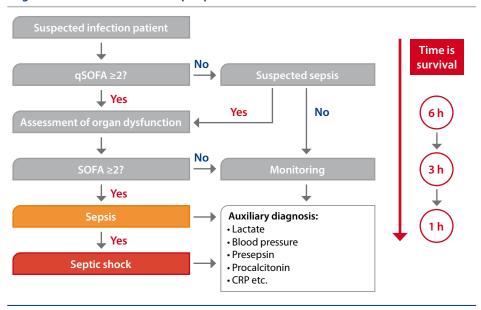


Fig. 3: Correlation of mortality rate caused by sepsis and time to effective antibiotic treatment



Adapted from World Sepsis Day by lindgruen-gmbh.com

Fig. 4: Scheme for detection of sepsis patients



qSOFA: quick Sequential Organ Failure Assessment, SOFA: Sequential Organ Failure Assessment, CRP: C-reactive protein; *Adapted from Singer et al.*, 2016 (15)

Medical scores and biomarkers for sepsis

Clinical scores like the Acute Physiology and Chronic Health Evaluation II (APACHE-II), Sequential Organ Failure Assessment (SOFA) score, Simplified Acute Physiology Score 2 (SAPS-2) are widely used scores in the ICU (16). The systemic inflammatory response syndrome (SIRS) criteria has been considered to be central to the diagnosis of sepsis, promoting the importance of inflammation for many years (17). SIRS criteria are composed of four symptoms (body temperature, respiratory rate, white blood cell and heart rate). When at least 2 out of 4 criteria are present, these patients are regarded as SIRS condition.

The third international consensus definitions for sepsis and septic shock (Sepsis-3) changed the definition of sepsis, where SOFA score (**Tab. 1**) should be used as the diagnostic criteria and quick SOFA (qSOFA) for non-ICU patients (3). qSOFA criteria (**Tab. 2**) shall be used as a warning system to draw attention to clinicians to perform further investigation for organ dysfunction (15).

Blood culture is considered as the most important test (the Gold standard) for the diagnosis of sepsis. However, it usually takes a day or more for microorganism cultivation and the positive blood cultures rate is not sufficient enough to determine the infection.

C-reactive protein (CRP), an acute phase protein increased at inflammatory states, is routinely used in patients with suspected infection. CRP however will be elevated for reasons other than bacterial infections such as burns, severe trauma and autoimmune diseases. Since formation of CRP is triggered by cytokines it will rise late (18).

Assicot et al. reported first in 1993 that Procalcitonin (PCT) specifically increased in bacterial infection (19). PCT becomes essential for diagnosis of sepsis and for guiding their therapy and monitoring (20). In 2016, the SSCC committee recommended to measure PCT for antimicrobial therapy guidance.

Next to CRP and PCT, many other biomarkers have already been evaluated at both hyper-inflammatory and immunosuppressive states like e.g. Leucocyte count, IL-6, IL-8, IL-10, PD-1/PDL-1, IL-1, CD64, TREM-1 and others (**Fig. 5**, 15, 21). Unfortunately, an ideal single biomarker has not yet been identified.

Tab. 1: Sequential (Sepsis-related) Organ Failure Assessment Score (SOFA-Score)

0,,,,,,	Ohioativa			Points		
Organ system	Objective - measurement	0	1	2	3	4
Respiration	PaO ₂ /FIO ₂	≥400	<400	<300	<200*	<100*
Coagulation	Platelets (cells/mm³)	>150,000	<150,000	<100,000	<50,000	<20,000
Liver	Bilirubin, mg/dL	<1.2	1.2 - 1.9	2.0 - 5.9	6.0 - 11.9	>12.0
Cardiovascular	MAP (mm Hg) or vasopressor	MAP ≥70	MAP <70	DPA ≤5	DPA 5.1 - 15	DPA >15
Central nervous system	GCS	15	13-14	10-12	6-9	3-6
Renal	Creatinine, mg/dL or urine output	<1.2 -	1.2-1.9 –	2.0-3.4 –	3.5-4.9 <500 mL/d	>5.0 <200 mL/d

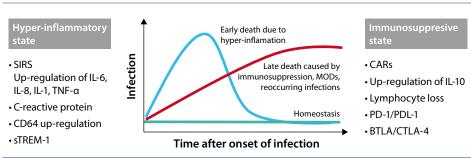
PaO₂: Partial pressure of oxygen (in arterial blood), FIO₂: Fraction of inspired oxygen, DAP: dopamine in mcg/kg/min for \geq 1 hour (note that SOFA also includes vasopressors other than dopamine in cardiovascular criteria), GCS: Glasgow Coma Scale score, MAP: mean arterial pressure; * With respiratory support; Adapted from Singer et al., 2016 (15)

Tab. 2: Quick SOFA (qSOFA) for diagnosis of suspected sepsis

Parameter	Criteria
Low blood pressure	SBP ≤100 mm Hg
High respiratory rate	≥22 breaths per min
Altered mentation	Glasgow coma scale <15

Adapted from Singer et al., 2016 (15)

Fig. 5: Sequential stages of sepsis



SIRS: Systemic Inflammatory Response Syndrome, IL: Interleukin, CD: Cluster of differentiation, sTREM1: Soluble triggering receptor expressed on myeloid cells-1, CARs: Chimeric antigen receptor, PD-1: Programmed cell death protein 1, PDL-1: Programmed Death Ligand 1, BTLA: B- and T-lymphocyte attenuator, CTLA-4: cytotoxic T-lymphocyte-associated Protein 4; Adapted from Singer et al., 2016 (15)

What is Presepsin?

sCD14-ST is a 13k Da fragment derived from cleavage of CD14, a glycoprotein of 55 kDa anchored to the membrane of monocytes, macrophages and polymorphic neutrophils. CD14 acts as a receptor for lipopolysaccharide (LPS) complexes and the specific LPS binding protein (LBP). It can bind to peptidoglycans and other surface structures present in both Gram-Positive and Gram-Negative bacteria. Once bound to the LPS-LBP complex, it activates the intracellular inflammatory response of the Toll-Like receptor 4 (TLR4)/MD2complex, triggering the host's inflammatory cascade against the infectious pathogenic agent. Phagocytosis and activity of plasma proteases (lysosomal enzymes, cathepsin D) result in the formation of the fragment sCD14 subtype, in particular the 13 kDa fragment of sCD14-ST known as Presepsin (Fig. 6; 22).

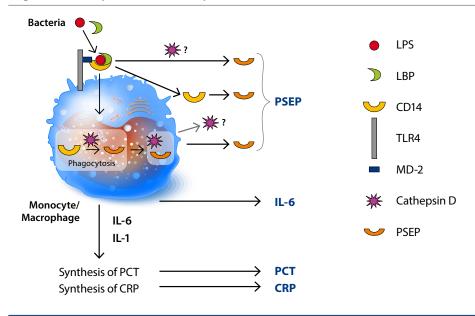
The half life of the molecule in plasma is 4-5 hours, compared to 12-24 hours for PCT, allowing more effective and earlier management of the pharmacological treatment (93). Presepsin was shown to be generated during the immune response. In sepsis patients, infected microorganisms are digested by the activities of monocytes or macrophages called phagocytosis. During phagocytosis, CD14 molecules are also digested by intracellular lysosomal enzymes, such as cathepsin D, result-ing in the fragmentation of CD14. The N-terminal CD14 fragment (Presepsin), is

circulated again into blood which was shown

by in vitro analysis (35, 36).

In addition to the phagocytosis- and receptor model, several hypotheses are proposed in terms of Presepsin production because of the findings that Presepsin was produced even in patients with a very low number of white blood cells (WBC). Presepsin is metabolized through the kidneys and excreted with the urine (36). So far, the biological activity of Presepsin remains unknown.

Fig. 6: Schematic production of Presepsin



TLR: Toll-like-receptor, LPS: Lipopolysaccharides, LBP: LPS-binding protein, MD-2: Molecular dynamic-2, PSEP: Presepsin, CD: Cluster of differentiation, PCT: Procalcitonin; Mod. from C. Chenevier-Gobeaux et al., 2015 (22)

Measurement of Presepsin

Presepsin can easily be measured from whole blood or plasma with the compact PATHFAST™ analyzer at the point of care or in the lab. The PATHFAST™ Presepsin immunoassay is based on chemiluminescence technology and shows excellent precision. No interference was observed from bilirubin, hemoglobin, lipids, triglyceride, or rheumatoid factors. Presepsin can be measured from 20 pg/mL to 20000 pg/ml without dilution (24). The fully automated procedure takes less than 17 minutes and it requires only 100 µl samples.



Low sample volume use is an ideal property especially for paediatrics where it is sometimes difficult to obtain a sufficient amount of blood from the neonates. In whole blood samples, the effect of hematocrit can be corrected automatically or manually. There is excellent correlation between whole blood and plasma results (24).

PATHFAST™ assays show no biotin interference since utilized monoclonal antibodies are alka-line phosphatase conjugated instead. Running a sample on PATHFAST™ is simple and does not require special skills.

12

In addition to Presepsin, the PATHFAST™ reagent menu offers several other STAT assays which can be used in sepsis diagnosis such as D-Dimer, NT-proBNP, hs-cTnl, CK-MB and hs-CRP. All assays are provided in economical precalibrated unit-use cartridges. Up to six samples can be tested in parallel in one single run. Whole blood samples should be measured within 4 hours after collection. Plasma can be stored in the refrigerator for 3 days or may be frozen. The stability of whole blood and plasma is shown in **Tab. 3**. Complete Blood Count (CBC) mixing leads to unspecificly increased Presepsin values, probably caused by shear forces, whereas mixing on roller mixers or inverting mixing does not increase PSEP values (25).



Specimens	Store condition	Stability	
Whole blood (Heparin, EDTA)	Room temperature	Within 4 hours	
Heparin-plasma	Refrigerator (2-8 °C)	Within 3 days	
	Freezer (<-20 °C)	One year	
EDTA-plasma	Refrigerator (2-8 °C)	Within 3 days	
	Freezer (<-20 °C)	One year	



How to interpret Presepsin values?

Okamura et al. reported the Presepsin values from the measurements of heparinized plasma and whole blood samples from 127 healthy volunteers. Presepsin concentrations ranged from 105-333 pg/ml in plasma and for whole blood, 98.3-314 pg/ml, respectively (23).

Another study reported the reference interval for Presepsin with overall reference limits of 55-184 pg/ml determined from two hundred individuals (120 females, median 39 years (18-75), which values were a little lower than the values previously reported.

No significant differences between males and females were shown. Also, no influence by age was shown (37). Several studies described higher reference ranges of neonates than those for adults (see neonatal section).

The accuracy of PATHFASTTM Presepsin was evaluated by Endo et al. in 2012. The cut off value of Presepsin for discrimination of bacterial and nonbacterial infectious diseases was determined to be 600 pg/ml, with clinical sensitivity and specificity of 87.8% and 81.4%, respectively. The area under the receiver ope-

rating characteristic curve (AUC) was 0.908 (38). Lu et al. compared biomarkers among sepsis, severe sepsis and septic shock patients. AUC values for Presepsin, PCT, CRP and WBC to differentiate sepsis from non-infectious SIRS were 0.954, 0.874, 0.859 and 0.723, respectively. The cut off of Presepsin for discrimination of sepsis and nonbacterial infectious SIRS was determined to be 407 pg/ml. (39). Yamamoto et al., evaluated Presepsin accuracy with the sepsis-3 definition criteria with the cut off value of 508 pg/mL (AUC = 0.88) (40).

Recently, several systematic reviews and metaanalysis reports describe specificity and sensitivity of Prespsin (41, 42, 43, 44, 45, 46, 47, 48). Kondo et al. included 18 studies from 12 counties and evaluated Presepsin diagnostic accuracy for sepsis as the AUC was 0.87 [95% CI 0.84 to 0.90], where the AUC of PCT was 0.84 [95% CI 0.81 to 0.87]. They concluded that the overall diagnostic performance of PCT and P-SEP for infection were comparable (**Tab. 4**; 48).

Tab. 4: Presepsin Meta-analysis

Author, year		Number of	Mean/		Sepsis	Cut off value		Prevalence	Sensitivity		Specificity	
		participants	median age	design	definition	PCT (ng/mL)	P-SEP (pg/mL)		PCT	P-SEP	PCT	P-SEP
Ali, 2016 (76)	Egypt	51	49.8	Prospective	Sepsis-3	0.85	907	0.647	60.6%	69.7%	88.9%	83.3%
Balci, 2003 (77)	Turkey	89	58	Prospective	Sepsis-1	2.415	-	0.461	85.4%	-	91.7%	-
Bauer, 2016 (78)	USA	219	59	Prospective	Sepsis-1	0.74	_	0.551	73.1%	_	74.2%	_
Behnes, 2014 (79)	Germany	116	62	Prospective	Sepsis-2	_	530	0.705	_	91.0%	_	53.6%
Çakir Madenci, 2014 (80)	Turkey	37	40	Prospective	ABA 2007*	0.759	542	0.393	75.4%	77.5%	78.7%	76.5%
Endo 2012 (30)	Japan	185	66	Prospective	Sepsis-2	0.5	600	0.622	86.1%	87.8%	78.6%	81.4%
Enguix-Armada 2016 (81)	Spain	388	63	Prospective	Sepsis-2	0.28	101.6	0.634	92.3%	81.7%	96.5%	96.5%
Gibot 2004 (82)	France	76	60	Prospective	Sepsis-1	0.6	-	0.618	83.0%	-	69.0%	-
Godnic 2015 (83)	Slovenia	47	N.A.	Retrospective	Sepsis-2	3.12	413	0.851	57.5%	85.0%	71.4%	57.1%
Klouche 2016 (62)	France	144	58	Prospective	Sepsis-1	0.5	466	0.694	80.0%	90.0%	59.1%	54.5%
Leli 2016 (84)	Italy	92	73	Prospective	Sepsis-1	4.4	843.5	0.281	84.0%	88.0%	84.4%	71.9%
Miglietta 2015 (85)	Italy	145	64.4	Retrospective	Sepsis-1	0.88	_	0.625	85.7%	_	83.3%	_
Romualdo 2014 (86)	Spain	226	67	Prospective	Original	0.45	729	0.164	75.7%	81.1%	64.0%	63.0%
Selberg 2000 (87)	Germany	33	47.9	Prospective	Sepsis-1	3.3	-	0.667	86.4%	-	54.5%	_
Takahashi 2016 (88)	Japan	103	68	Prospective	Sepsis-1	0.85	658	0.85	78.8%	72.9%	73.3%	60.0%
Ugarte 1999 (89)	Belgium	190	62	Prospective	Sepsis-1	0.6	_	0.584	67.6%	_	60.8%	_
van der Geest 2016 (90)	Netherlands	301	57	Prospective	Original	1.41	_	0.505	65.1%	_	66.4%	_
Wong 2013 (91)	France	270	61	Prospective	Not described	0.5	_	0.537	88.3%	_	64.0%	_
Yang 2016 (92)	China	300	64	Prospective	Sepsis-1	0.4475	_	0.357	83.2%	_	53.9%	_

PCT: Procalcitonin, P-SEP: Presepsin; *American Burn Association Consensus Criteria; Adapted from Kondo et al., 2019 (51)

In clinical studies performed in Peru and Germany, cut off values were established. For healthy individuals the normal values of Presepsin are below 200 pg/mL. A Presepsin cut off value of 622 pg/ml excludes 30 day mortality by a Negative Predictive Value (NPV) of 98.5% (28). Based on the Presepsin values measured in the study patients with different disease severity degrees (SIRS, sepsis, severe sepsis or septic shock) and the close relationship between Presepsin and outcome decision thresholds for risk stratification could be established (**Tab. 5**: 22, 26).

In Germany, another study examined the diagnostic and prognostic validity of Presepsin in 140 emergency patients. Similar decision thresholds could be identified for diagnostic differentiation of sepsis grades and mortality prediction in septic patients presenting at the Emergency Room (ER). The determination of Presepsin at presentation allowed reliable risk stratification already at the earliest time point in patients suspected for sepsis. Moreover, the Presepsin concentration during anti-microbial therapy was related to the patient's outcome (**Tab. 6**: 27).

Recommendation

For a risk assesment in adult patients, threshold values given in **Tab. 5 and 6** may be used. For final diagnosis, Presepsin results may support clinical findings but should not be used as a sole decision criteria.

16

Tab. 5: Diagnosis of sepsis by Presepsin

Presepsin (pg/mL)	Diagnosis
< 200	Exclusion of sepsis
200-300	Systemic infection not probable
300-500	Systemic infection (sepsis) possible
500-1000	Significant risk of the systemic infection progression (severe sepsis), increasing risk of unfavorable outcome
≥1000	High risk of the systemic infection progression (severe sepsis/septic shock). High risk for mortality after 30 day comparable with a SOFA score ≥ 8

Adapted from Carpio et al., 2015 and Chenevier-Gobeaux, 2015 (22,26)

Tab. 6: Presepsin decision thresholds at admission of 30 days outcome

Risk stratification	Very low	Low	Moderate	High	Very high
Presepsin (pg/mL)	<200	200-300	300-500	500-1000	≥1000
Low grade sepsis, n (%)	3 (3.5)	9 (10.6)	18 (21.1)	29 (34.1)	26 (30.6)
Severe sepsis, n (%)	0	0	5 (12.5)	11 (27.5)	24 (60.0)
Septic shock, n (%)	0	0	0	4 (26.7)	11 (73.3)
30-day death, n (%)	0	0	0	5 (21.7)	18 (78.3)

Adapted from Spanuth et al., 2011 (27)

Specificity and sensitivity of Presepsin

Early sepsis marker

One of the Presepsin advantages is the quick response against the onset of sepsis. A case study at Iwate Medical University compared the sepsis biomarkers Presepsin, PCT and CRP after a colon perforation with septic shock.

Presepsin showed a high concentration level spontaneously at the infectious event and decreased in parallel with the APACHE II score. On the other hand, IL-6, CRP and Procalcitonin elevated one day after the onset of disease (**Fig. 7**).

Low influence from burns, trauma and medical surgery

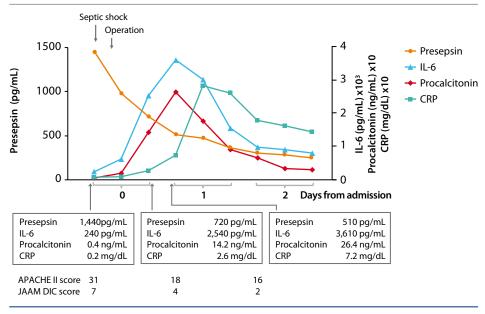
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Because CRP and PCT are inflammatory biomarkers, it is well known they are affected by burns, severe trauma, or surgical operations other than the sepsis status. Presepsin values are less affected by such cases. A typical case report of a severe burn patient indicated that IL-6, CRP and PCT increased in the first period (day 0 to 5) due to burn inflammation whereas Presepsin did not. When this patient became septic, Presepsin increased earlier than the other markers and showed a peak at the septic shock. After polymyxin B-immobilized fiber

column (PMX-DHP) therapy on day 8 Presepsin decreased as expected whereas IL-6, PCT and CRP decreased delayed and did not reflect clinical course correctly (**Fig. 8**).

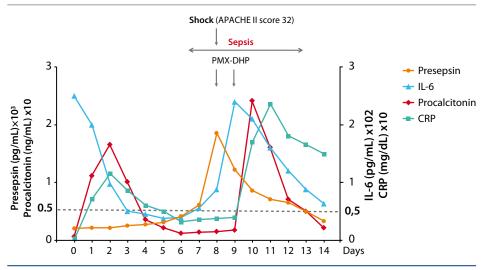
Hoshino et al. also showed, in severe trauma patients with Injury Severity Scores ≥16, a significant increase of PCT on day 1 after trauma while Presepsin concentration was neither elevated at day 0 nor day 1. This result strongly indicated that Presepsin was useful to diagnose sepsis in severe trauma patients (28).

Fig. 7: Behavior of sepsis biomarker in a colon perforation patient



IL: Interleukin, CRP: C-reactive protein; Data kindly provided by Dr. Endo, Iwate Medical University

Fig. 8: Behavior of sepsis biomarker in a burn patient who received PMX-DHP



PMX-DHP: hemoperfusion using polymyxin B-immobilized fiber column, IL: Interleukin, CRP: C-reactive protein; Data kindly provided by Dr. Endo, Iwate Medical University

In the case study by Shozushima et al., a 51-year-old patient was taken to hospital with extensive burns covering 76% of his body. The laboratory data on arrival included an elevated white blood cell (WBC) count of 38,880/uL, and a diagnosis of SIRS was made. Since no elevation of the Presepsin value (281 pg/ml) or PCT value (0.98 ng/ml) was observed on arrival and the blood cultures were negative, there was no suspicion of sepsis at that time.

Presepsin (in orange) and other markers in post-traumatic patients following a serious burn were considered. It is well-known that Presepsin values does not change after the trauma and there is an early increase in the values of Presepsin by day 2 following the occurrence of a bacterial infection confirmed by a positive blood culture of day 5. Effectiveness of anti-biotic treatment is shown at day 13 (**Fig. 9**, 24).

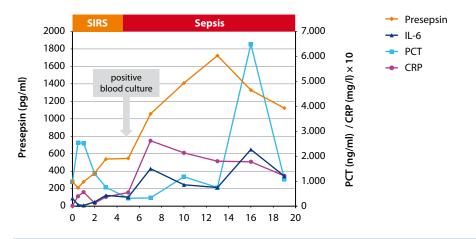
Prognosis and monitoring with Presepsin

Evolution of Presepsin levels over time in survivors was significantly different from that in deceased patients in the ICU. PCT levels decreased rapidly and similarly in both survivors and non-survivors whereas Presepsin clearly differentiates already after 24 hours between the two cases.

In comparison to survivors, Presepsin levels in non-survivors stayed constantly high over the time period observed. Conversely, PCT levels fell rapidly and similarly from day 1 to 7 in survivors and non-survivors. Presepsin appears as an early marker of mortality with better prognostic performance than PCT and can be used as an aid in risk stratification strategies in septic patients (**Fig. 10**; 29).

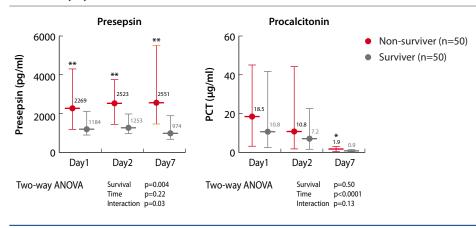
Patients with decreasing levels of Presepsin over 7 days in ICU were more likely to have received an early appropriate first-line empirical antibiotic therapy on day 1 than those with increasing levels (29).

Fig. 9: Case report of sepsis biomarker in a burn patient



IL: Interleukin, CRP: C-reactive protein, PCT: Procalcitonin; Adapted from Shozushima et al., 2011 (24)

Fig. 10: Time course of plasma concentrations of Presepsin and Procalcitonin during ICU stay by survival status



Adapted from Masson et al., 2014 (29)

In a multicentric study in Japan with 140 patients observed over a 7 day period, Presepsin and other biomarkers used in sepsis were investigated in sepsis patients over the clinical course. All markers declined over time in patients with predicted favorable outcome according to SOFA or APACHE II scores. Unlike other biomarkers, only Presepsin values showed a tendency to stay elevated in the group of patients with unfavorable outcomes.

A clear difference in the development of Presepsin and PCT values during the course of treatment could be demonstrated with sepsis patients who got antimicrobial treatment after diagnosis of sepsis. Presepsin showed a clear trend towards lower values in survivors over the period from 0-72 h observation time while non-survivors reached very high values. PCT, in contrast, though also much higher in non-survivors, showed only a marginal decline after 24 hours in the survivors (**Fig. 11**; 30).

Recommendation

For ICU patients, a baseline cut off for Presepsin of approximately 1,000 pg/ml may be used which may increase with progression of disease and severity of organ failure.

For final diagnosis, Presepsin results may support clinical findings but should not be used as a sole decision criteria for severity of organ damage.

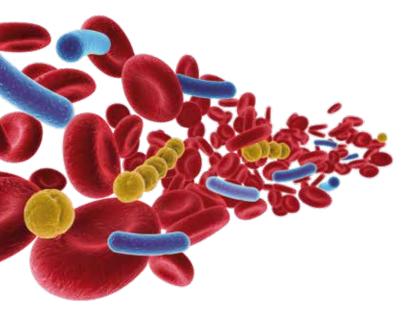
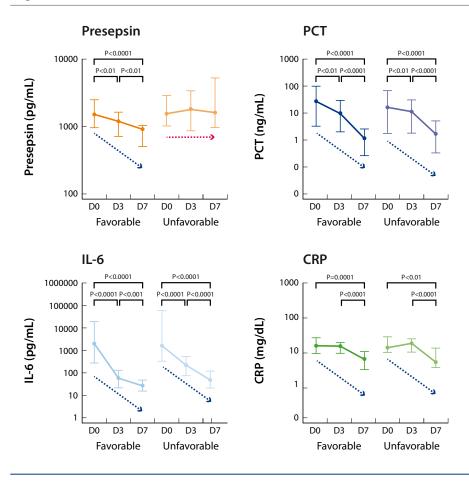


Fig. 11: Course of biomarkers over time



IL: Interleukin, CRP: C-reactive protein, PCT: Procalcitonin; Adapted from Endo et al., 2014 (30)

Organ dysfunction and Presepsin

According to the Sepsis 3 definition, organ dysfunctions are main symptoms of sepsis which indicates that Presepsin concentration reflects the severity of sepsis.

Masson S et al. demonstrated the association of Presepsin concentration at baseline with the incidence of new organ failures. Presepsin concentration at baseline (946 ng/L) increased with the SOFA score, the number of prevalent organ dysfunctions or failures, and

Presepsin is metabolized rapidly through the kidneys and was reported to be affected by kidney failure. Nagata et al. studied Presepsin values in patients with various chronic kidney disease (CKD) stages. Even if patients were not infected by any pathogens, Presepsin values were increased by the decrease of glomerular filtration rate (GFR) values. Moreover, Presepsin values elevated extensively among the patients who received hemodialysis (HD), so other cut off values need to be used (32).

Another study reported the median of Presepsin in hemodialysis treated and that the optimal cut off value was 2,083 pg/ml (33). Acute kidney injury (AKI) is one of the major complications in sepsis patients. Takahashi et al. showed that the diagnostic accuracy for infec-

the incidence of new respiratory, coagulation, liver, cardiovascular and kidney system events (**Fig. 12,** 31). The concentration decreased over 7 days in ICU patients with negative blood cultures. Presepsin increased with inappropriate antibiotic therapy. Baseline Presepsin was independently associated and correctly reclassified with the risk of ICU and 90-day mortality (31).

tious diseases in patients with AKI was even higher than that in patients without AKI, however, the cut off value for diagnosis of sepsis among AKI patients should be changed. Plasma levels of neutrophil gelatinase-associated lipocalin (NGAL), a kidney dysfunction marker, was used to be classified into non-AKI (<150 ng/mL) and AKI (≥150 ng/mL) groups. AUCs of Presepsin were higher than those of PCT in non-AKI and AKI groups (AKI group PSEP 0.83 vs. PCT 0.72 / non-AKI group PSEP 0.75 vs. PCT 0.67). The optimal cut off values of Presepsin for AKI and Non-AKI patients were 828 pg/mL and 694 pg/mL, respectively (Fig. 13, 34). Presepsin seems to be a useful biomarker for bacterial infections in AKI patients but diffe-

Recommendation

AKI patients without infections and Non-AKI patients with infections show similar cut off values (about 600 pg/ml) which reflects the influence of the organ damage to the Presepsin level. When AKI patients get an additional infection, cut off values can even rise up to 1,200 pg/ml, whereas Non-AKI

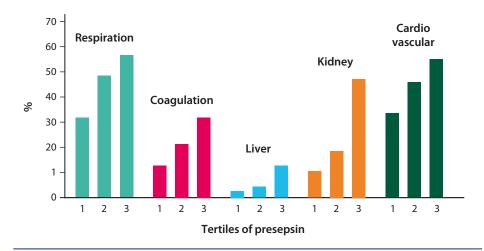
patients without infection show normal cut off values of approximately 300 pg/ml (see also **Tab. 5 and 6**).

rent thresholds should be applied (34).

For final diagnosis, Presepsin results may support clinical findings but should not be used as a sole decision criteria severity of organ damage.

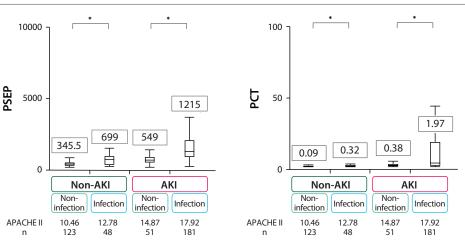
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Fig. 12: Baseline Presepsin concentration according to the incidence of new organ failures



Adapted from Masson et al., 2015 (33)

Fig. 13: Comparison of levels of PCT and Presepsin between non-infection and infection groups in non-AKI and AKI patients using NGAL



Median values are indicated in the boxes; *P < 0.01; PCT: Procalcitonin, NGAL: Neutrophil Gelatinase-Associated Lipocalin, AKI: Acute Kidney Injury; Adapted from Takahashi et al., 2016 (34)

Pathogens and Presepsin

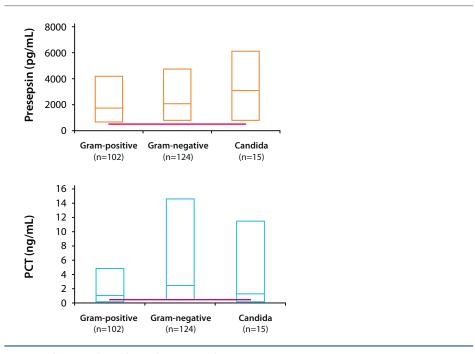
Due to the Presepsin production mechanism build up from CD14 by the mechanism of phagocytosis, the specificity of microorganisms was investigated. Endo et al. initially reported that there were no significant differences in Presepsin levels between the Gram-positive and Gram-negative bacterial infection groups (2,881 \pm 4,374 and 2,641 \pm 3,709 pg/ml, respectively; P = 0.5320) (38) and several other reports support this result. Rabensteiner J et al., examined 300 patients consecutively included 100 for Gram-positive and Gram-negative bacteremia, 50 for candidemia and 50 for controls. The median of Presepsin of Gram-positive, Gram-negative and candemia were 1078, 1295 and 2293 pg/mL, respectively (Fig. 14; 49).

Ugajin et al., examined the Presepsin levels in pneumonia patients and revealed that Presepsin levels were different between Grampositive and Gram-negative bacteria. These differences may be related to the difference of species, or sepsis severity (50).

Qi et al. examined the usefulness of differentiating active pulmonary tuberculosis (APTB) from bacterial community acquired pneumonia (BCAP). Presepsin concentrations in APTB patients were slightly increased, and may be helpful for initial discrimination between APTB and BCAP. This is probably caused by PSEP levels and the low immunogenicity by Mycobacterium tuberculosis (52).

There is little information about virus infection. Arai et al. analyzed for a cohort study of patients undergoing allogeneic hematopoietic cell transplantation and reported that patients with hemophagocytic syndrome (HPS) and bacteremia (n=19) showed higher levels of Presepsin. No correlation between Presepsin and CMV (cytomegalovirus) reactivation (n=12) could be shown (53). Moreover, a study with febrile children showed that Presepsin values in patients with Influenza A did not elevate in contrast to other bacterial infections or Kawasaki diseases (54).

Fig. 14: Levels of Presepsin and Procalcitonin in Gram-positive bacteria, Gram-negative bacteria and candida



PCT: Procalcitonin; Adapted from Rabensteiner et al., 2014 (49)

Presepsin and invasive fungal infections

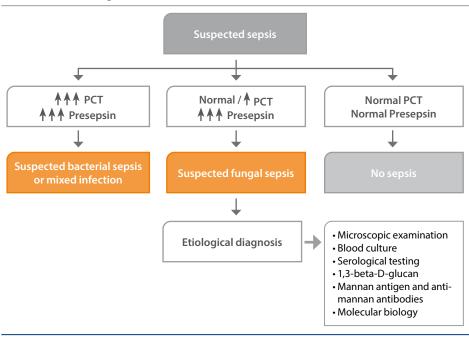
Lippi et al. investigated the usefulness of Presepsin and Procalcitonin in the context of invasive fungal infections. Invasive fungal infections are a major healthcare issue accounting for approximately 20% of all sepsis cases. Classic fungal diagnostics are serologic testing, measurement of 1,3-beta-D-glucan, mannan antigen or anti-mannan antibodies and molecular biology. These findings lead the way to developing diagnostic algorithms based on results of both biomarkers Procalcitonin and Presepsin.

Concomitantly increased values of these biomarkers could be suggestive of bacterial sepsis or mixed infection. Non-diagnostic values of both biomarkers may enable the ruling out of sepsis of bacterial or fungal origin. A disproportionate increase of Presepsin values combined with normal or only modestly elevated Procalcitonin concentration may be suggestive of invasive fungal infections (**Fig. 15**; 55).

Moreover, Bamba et al. evaluated cases of fungal bloodstream infections and found elevated Presepsin levels correlated with increased SOFA score. They also showed that Presepsin increased by incubation with blood and Candida sp. in vitro (51).



Fig. 15: Algorithm based on Procalcitonin (PCT) and Presepsin test results for screening invasive fungal infections



PCT: Procalcitonin; Adapted from Lippi et al., 2019 (55)

Application of Presepsin in a clinical setting

Since the first publications of clinical use of Presepsin in 2005, about 240 papers from various categories (as of June/2019), such as ED (47 papers), ICU (22 papers), hematology (15 papers), neonates (24 papers) and others were published during the last 14 years (**Fig. 16**).

Other than sepsis, clinicians are interested in Presepsin in terms of specificity for bacterial infection including local infection. Due to the high interest in the clinical use of Presepsin, more publications will follow.

Neutropenia

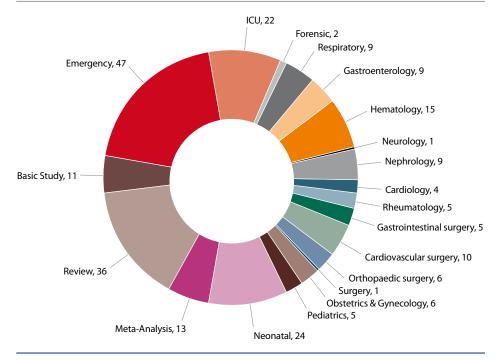
Febrile neutropenia (FN) remains one of the major complications during chemotherapy in patients with solid tumors or hematological malignancies. FN is basically defined as having an oral temperature ≥38.3 or ≥38°C for more than 1 h with absolute neutrophil count (ANC) <500 cells/µL or the reduction to 500 cell/µL in the next 24-48 h. The mortality rate is higher and it is difficult to predict or to detect those mali-cious events at an earlier point in time. Koizumi et al. showed Presepsin values in FN patients were not significantly associated with the absolute WBC count and the median plasma Presepsin level was 456.5 pg/mL even with a low WBC count below 100/µL. This data revealed that Presepsin values were increased in the case of FN in spite of the Presepsin production mechanism. One case study described a

48 year old male patient with B-cell leucaemia who received chemotherapy. On day 10 he experienced infection with K. pneumonia. Plasma Presepsin level was already elevated one day prior to FN onset whereas CRP was not increased at that time (**Fig. 17**; 56)

Ebisawa et al. studied Presepsin and Procalcitonin response time after the onset of fever, and showed that Presepsin has an advantage for early detection (from 1 to 18 hours) with the AUC of 0.8188, where PCT has little elevation (AUC=0.6354) (57).

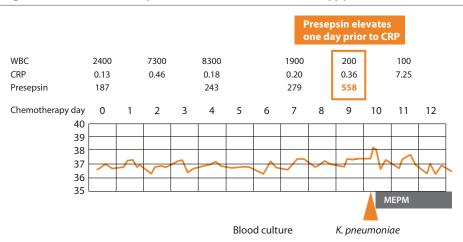
Nanno et al. reported higher median values of Presepsin (>1935 pg/mL) with hemaphagocytic syndrome (HPS) and Presepsin values were significantly associated with the 90-day mortality (58).

Fig. 16: Applications of the biomarker Presepsin



Source: Pubmed and Google Scholar

Fig. 17: Time course of Presepsin, CRP and WBC in a chemotherapy patient



WBC: White Blood Cells, CRP: C-reactive protein, MEPM: Meropenem; Adapted from Koizumi et al., 2017 (56)

Perioperative risk assessment

Differentiation between SIRS and sepsis in surgical patients is of crucial significance. Inflammatory biomarkers are commonly measured in clinical practice, however, CRP and PCT are influenced by non-specific systemic inflammatory response (24). Presepsin is expected to be specific for bacterial infection at periopera-

tive periods because of less response against burns and trauma (23). Takahashi G. presented that in comparison to CRP and WBC, Presepsin shows no unspecific increase via surgical interventions in the case of spinal scoliosis surgery (personell communication) (n=12) (**Fig. 18**).

Popov et al. evaluated the prognostic values of Presepsin in cardiac surgery. During study periods 19 out of 51 patients (37%) developed sepsis in which the most frequent complication was ventilator-associated pneumonia (n=12). Presepsin values showed significant differences between with and without infectious complications during the first day after the operation. Presepsin dynamics in the postoperative period were found with an increase of Presepsin levels associated with higher risk of infection. When Presepsin levels were persistently higher than the normal values, more than 50% of patients (21/24 58.3%) acquired infectious complications (59). Another study from Germany also revealed high prognostic value in cardiac surgery patients. 856 patients were studied prospectively. Preoperative plasma concentrations of Presepsin, Procalcitonin, NT-proBNP, Cystatin C and the the additive

> initial empiric antibiotic therapy, where Presepsin values decreased to the threshold level at T3 (144h after operation). Four patients who died due to sepsis showed increased Presepsin values within 28 days from admission (61).

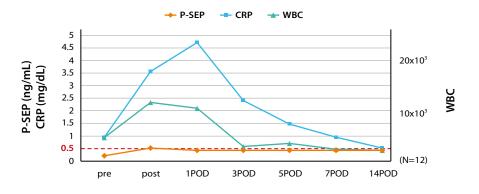
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A researcher group from Italy recruited 35 cadaveric organ transplant recipients and 35 abdominal surgery patients and measured Presepsin after surgery. Presepsin levels were very high in the case of blood culture positive group (n=50) and 33 patients which received

European System of Cardiac Operative Risk Evaluation 2 were compared to mortality at 30 days, 6 months and 2 years.

Thirty-day mortality was 3.2%, 6-month mortality was 6.1%, and 2-years mortality was 10.4% across the population. The median of pre-operative Presepsin concentrations was significantly higher in 30-day nonsurvivors than in survivors: 842 pg/ml versus 160 pg/ml difference. The results were similar for 6-month and 2-years mortality. Presepsin also provided better discrimination than Cystatin C, N-terminal pro-hormone natriuretic peptide, or Procalcitonin. An elevated preoperative plasma Presepsin concentration is an independent predictor of postoperative mortality in elective cardiac surgery patients and is a stronger predictor than several other commonly used biomarkers and scores (Tab. 7; 60).

Fig. 18: Presepsin is not influenced by surgical trauma



WBC: White Blood Cells, CRP: C-reactive protein, POD: Post Operative Day;
Data kindly provided by Dr. Endo and Dr. Takahashi

Tab. 7: Comparison of Presepsin with further risk factors

		ROC analysi	s to predic	t 30-day	mortality (n=	=852)	P-value vs. AUC
	AUC	95% CI	P-value	Cut off	Sensitivity, %	Specificity, %	of preoperative Presepsin
Preoperative							
Presepsin, mg/dL	0.88	0.81-0.96	< 0.001	>295	84	83	-
Leukocytes, ×10 ⁹ /L	0.58	0.45-0.71	0.18	≥11.1	28	93	<0.001
Procalcitonin, ng/ml	0.59	0.48-0.69	0.13	≥0.022	56	68	<0.001
Age, years	0.73	0.64-0.81	< 0.001	≥ 67.3	84	51	0.004
EuroSCORE 2	0.74	0.65-0.83	< 0.001	>4.4	84	57	0.02
NT-proBNP, pg/mL	0.77	0.69-0.84	< 0.001	>676	96	57	<0.001
Cystatin C, mg/dL	0.76	0.64-0.87	< 0.001	>1.65	64	87	0.02
Intraoperative							
Operation time, min	0.76	0.66-0.87	< 0.001	>206	64	78	0.06
Postoperative							
Procalcitonin, ng/mL	0.85	0.76-0.94	< 0.001	≥2.0	84	76	0.51
Presepsin, pg/mL	0.85	0.77-0.93	< 0.001	>726	76	86	0.15

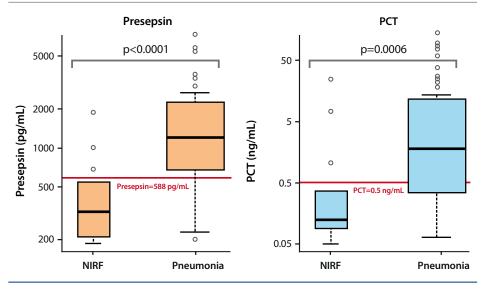
Adapted from Bomberg et al., 2017 (60)

Pneumonia

Some reports show the usefulness of Presepsin measurement for pneumonia patients (62, 50, 52, 73, 74). Klouche et al. pointed out that among ICU-admitted patients, Presepsin values in severe pneumonia group were significantly higher than in non-infectious group, and Presepsin reflected prognosis as well (**Fig. 19**; 62). Ugajin et al. also showed that Presepsin values at admission predicted 30-day morality rate using cut off 470 pg/ml (**Fig. 20**; 50).

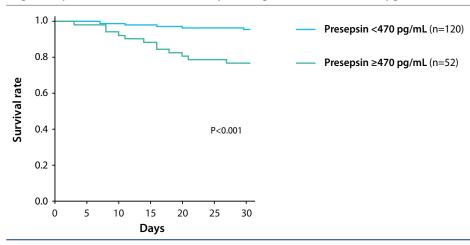


Fig.19: Comparison of Presepsin and PCT levels for non-infectious and pneumonia patients



NIRF: Non-infectious respiratory failure; Adapted from Klouche et al., 2016 (62)

Fig. 20: Kaplan-Meier survival curve Presepsin using the cut off value of 470 pg/ml



Adapted from Ugajin et al., 2019 (50)

Other infectious diseases including local infections

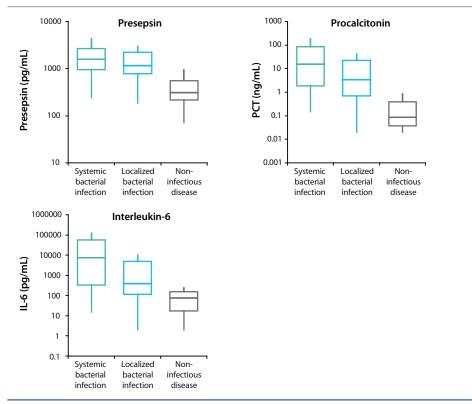
Endo et al. showed in a multicenter prospective study (n=207) that Presepsin could be produced even by local infection. The cut off value of Presepsin for discrimination of bacterial and nonbacterial infectious diseases was determined to be 600 pg/ml, of which the clinical sensitivity and specificity were 87.8% and 81.4%, respectively. The sensitivity of blood culture was 35.4%; that for Presepsin was 91.9%. Moreover, there were no significant differences in Presepsin levels between the blood culture-positive and -negative groups (**Fig. 21**; 38).

Salina et al. (95) from a study with acute pancreatitis (SAP) patients and septic complications point to Presepsin as an early marker for supporting diagnosis of SAP.

Lin et al. showed the relationship between the severity of acute cholangitis (AC) and Presepsin values. 119 AC patients were classified from Grade I to III using the TG13 criteria, Presepsin values were increased with severity (**Fig. 22**; 63).

36

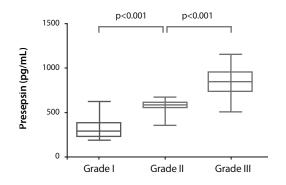
Fig. 21: Distribution of Presepsin, PCT and IL-6 in systemic infections, localized bacterial infections and non-infectious disease



IL: Interleukin; Adapted from Endo et al., 2012 (38)

Fig. 22: Box plot of Presepsin concentration according to the severity of acute cholangitis

37



Adapted from Lin et al., 2016 (63)

Imagama et al. studied the usefulness of Presepsin for the identification of septic arthiritis (SA) from osteoarthritis (OA). The mean blood level of Presepsin was 529.4 ± 470.8 pg/ml in

the SA group and 136.4 ± 52.4 pg/ml in the OA group. Interestingly, the diagnostic accuracy was significantly improved using synovial fluid (**Fig. 23**; 64).

Another scientific report provided information regarding different causes of pleural effusions. Presepsin values were significantly higher in the case of empyema, which was caused by infection (65). Presepsin values are higher in pleural fluids than in blood, strongly suggesting that Presepsin was produced at infectious loci (26).

Titova et al (96) found in hemodialysed patients with pneumonia and sepsis three times higher levels of Presepsin compared to other hemodialysed patients w/o sepsis.

Abudeev et al. (94) reported Presepsin use in cerebrospinal fluids (CSF) to support diagnosis of nosocomial infections of the central nervous system.

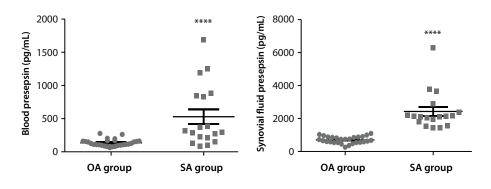
Shiota et al. reported (33) severe local infection during critical limb ischemia (CLI) patients who received hemodialysis. As mentioned above, hemodialysis affects the Presepsin baseline; however, Presepsin values were significantly increased in non-healing group where bacterial infections were strongly suspected. This study indicated the possibility to detect infection even when receiving hemodialysis (**Fig. 24**; 33).

Other diseases that might affect Presepsin levels

Presepsin has also been studied in other cases than sepsis or infectious diseases. Tanimura et al. researched the clinical significance of Presepsin levels in patients with systemic lupus erythematosus (SLE). Elevated plasma Presepsin levels were correlated with disease activity of SLE, suggesting inappropriate monocyte or neutrophil activation in the pathophysiology of SLE exacerbation (66). Moreover, Presepsin was linked to early identification of severe acute pancreatitis (SAP). Presepsin levels showed obviously higher levels in SAP patients than in healthy individuals (67). Acute myocardial infarction could also be also considered to be a state of inflammation with activated monocytes.

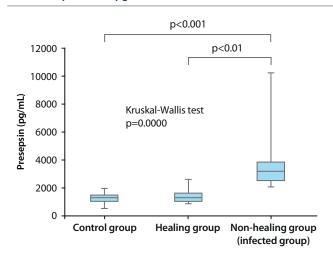
Caglar et al. examined Presepsin levels in patients with acute ST elevation myocardial infarction (STEMI). Plasma Presepsin and troponin levels were significantly higher in patients with STEMI than controls (1988.89 \pm 3101.55 vs. 914.22 \pm 911.35 pg/ml, p = 0.001 and 3.46 \pm 3.39 vs. 0.08 \pm 0.43 ng/ml, p = 0.001, respectively). The cut off value for Presepsin of 447 pg/ml was found to detect STEMI with 87.5% sensitivity, 44% specificity, 60% positive predictive value and 78.5% negative predictive value. Presepsin levels were found to be significantly elevated in patients with STEMI together with high-sensitivity troponins (68).

Fig. 23: A comparison of the concentrations of synovial fluid Presepsin between septic arthritis and osteoarthritis



OA: osteoarthritis, SA: septic arthritis; Adapted from Imagama et al., 2019 (64)

Fig. 24: Box plot of the plasma Presepsin concentrations in the control group, healing group, and non-healing group. The median value is indicated in the center of the box, and is expressed in pg/mL



Adapted from Shiota et al., 2016 (33)

Neonatals

Sepsis is one of the most significant syndromes for mortality and morbidity in the neonatal population. The diagnostic validity of Presepsin in neonatal sepsis has been evaluated in numerous clinical studies.

Pugni et al. enrolled 684 healthy neonates for evaluation of reference ranges for Presepsin. The Presepsin median value in term infants was 604 pg/mL whereas, in preterm infants the Presepsin median value was slightly higher (620 pg/mL). The normal reference ranges of Presepsin observed were higher than those seen in healthy adults (69).

Generally, neonatal sepsis is classified either as early onset sepsis (EOS) occurring in the first 72 h of life or late onset sepsis (LOS) which occurs at day 4 or after birth.

Poggi et al. prospectively studied preterm newborns (≤32 weeks gestational age) with

Recently, two meta-analysis for the diagnostic accuracy of Presepsin in neonatal sepsis were published. Bellos et al. selected 11 clinical study publications with a total number of 783 neonates. The pooled sensitivity of Presepsin for the prediction of neonatal sepsis was 0.91, the pooled specificity was 0.91 and the diagnostic odds ratio was 170.28 (95% Cl 51.13-567.11). Head-to-head comparison with AUC values of CRP (0.9748 vs. 0.8580) and PCT (0.9596 vs. 0.7831) revealed that Presepsin was

LOS and non-infected controls. Presepsin was higher in the LOS than in the control group at enrollment (1295 vs. 562 pg/mL) throughout the evaluation period. The area under the ROC curve was 0.972. P-SEP achieved the best accuracy for prediction of probable sepsis at the cut off of 885 ng/L (**Fig. 25;** 70).

Montaldo et al. studied preterm neonates (<34 wk. gestational age) who were admitted to NICU by 6 hours of age with suspicion of sepsis. Presepsin values are significantly higher in the EOS group than those in the uninfected group at every time interval, and the highest accuracy was achieved at 24 h after birth with the AUC score 0.97 using the cut off value of 788 pg/ml. Importantly, CRP as well as PCT are influenced by the physiological change during the first day of life, so cut off points should be determined in a time-dependent manner (71).

more accurate in detecting neonatal sepsis. The cut off values from each publication were categorized into 650, 650-850, 850 pg/ml as listed in **Tab. 8** which indicated that the diagnostic efficacy was maximized (AUC 0.99) when the Presepsin cut off value in neonates ranged from 650-850 pg/ml (72). Ruan et al. (75) also showed superior diagnostic accuracy of Presepsin for neonatal sepsis diagnosis when compared with simultaneous use of CRP and PCT (AUC = 0.99 vs AUC = 0.96) (75).

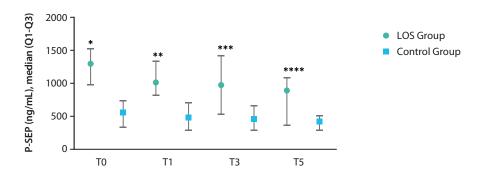
Recommendation

A preliminary cut off value of ≤650 pg/ml for exclusion of sepsis seems to be appropriate for neonates. Presepsin values ≥850 pg/ml indicate a bacterial infection.

For final diagnosis, Presepsin results may support clinical findings but should not be used as a sole decision criteria for severity of organ damage.

40

Fig. 25: Change and comparison of Presepsin values in infants in the LOS and control groups



LOS: Late Onset Neonative Sepsis; Adapted from Poggi et al., 2017 (70)

Tab. 8: Diagnostic efficacy of Presepsin with pre-defined cut offs

Cut off (pg/ml)	Number of studies	Sensitivity (95% CI)	Specificity (95% CI)	DOR (95% CI)	AUC (SE)
≤ 650	3	0.91 (0.83–0.96)	0.85 (0.77–0.91)	71.78 (7.46–690.56)	0.9634 (0.0439)
650-850	5	0.91 (0.86–0.95)	0.97 (0.94–0.99)	542.72 (156.62–1880.60)	0.9915 (0.0046)
≥ 850	3	0.90 (0.82–0.95)	0.86 (0.77–0.93)	75.60 (8.32–686.53)	0.9681 (0.0316)

DOR: Diagnostics Odds Ratio, AUC: Area Under The Curve; Adapted from Bellos et al., 2018 (72)



Conclusions

The major advantage of the assessment of Presepsin is it's capacity to predict the severity of a bacterial infection. Presepsin correlates significantly with the degree of severity of the infection as its quantitative results increase proportionally. In fact, the studies reveal maximum correlation with the SOFA score values (clinical scoring used most frequently to evaluate organ failure). Higher values on the first day of monitoring are closely associated with a higher incidence of new organ failure and hemodynamic instability in the first 24 hours.

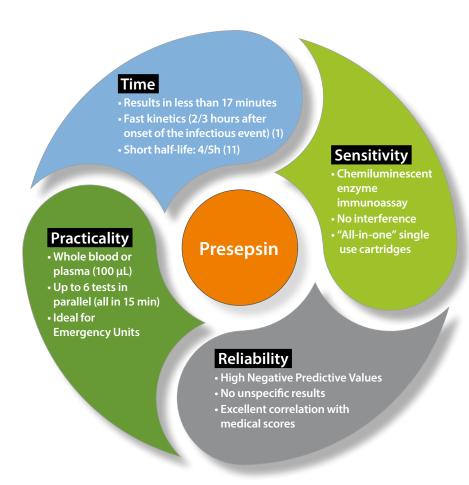
Presepsin concentration increased with the SOFA score, the number of prevalent organ dysfunctions or failures, and the incidence of new failures of the respiratory, coagulation, liver, and kidney systems. Presepsin is an early predictor of host response and mortality in septic patients. Changes in concentrations over time reflects the appropriateness of antibiotic therapy.

In addition, the measurement of Presepsin can be done by an easy procedure that takes less than 17 min with PATHFAST™.

Since only one single biomarker is not enough for a reliable diagnosis or prognosis of disease, support of Presepsin determination could be used in combination with other biomarkers and standard methods of infection diagnosis like e.g. medical scores for increased accuracy.

The different performance efficiency values may be due to the heterogeneity established in all the different studies. Possible factors for heterogeneity might be study strategy (prospective or not), clinical setting (ED, ICU), type of patients (adults or neonate), reference for sepsis criteria and even the type of sample (plasma or whole blood) for measurement of Presepsin. The better knowledge of conditions, that influence the levels of Presepsin, might enable reducing the false positive rate of infection diagnosis and inappropriate treatments. Recommended cut off values reflect the up-to-date-information from clinical studies and may be used as an orientation.

Future studies are necessary, for the identification of these conditions and the determination of cut off values for the detection of different types of infections in different groups of patients would also be effective in the clinical application of this biomarker.



Literature

- C. Fleischmann, A. Scherag, N. K. J. Adhikari, C. S. Hartog, T. Tsaganos, P. Schlattmann, D. C. Angus and K. Reinhart, "Assessment of Global Incidence and Mortality of Hospital-treated Sepsis. Current Estimates and Limitations," American journal of respiratory and critical care medicine, vol. 193, no. 3, p. 259–272, 2016.
- C. M. Torio and B. J. Moore, Healthcare Cost and Utilization Project (HCUP) Statistical Briefs, Rockville (MD), 2006.
- C. W. Seymour, T. D. Rea, J. M. Kahn, A. J. Walkey, D. M. Yealy and D. C. Angus, "Severe sepsis in prehospital emergency care: analysis of incidence, care, and outcome," American journal of respiratory and critical care medicine, vol. 186, no. 12, p. 1264–1271, 2012.
- C. J. Paoli, M. A. Reynolds, M. Sinha, M. Gitlin and E. Crouser, "Epidemiology and Costs of Sepsis in the United States-An Analysis Based on Timing of Diagnosis and Severity Level," Critical care medicine, vol. 46, no. 12, p. 1889–1897, 2018.
- C. Fleischmann-Struzek, D. M. Goldfarb, P. Schlattmann, L. J. Schlapbach, K. Reinhart and N. Kissoon, "The global burden of paediatric and neonatal sepsis: a systematic review," The Lancet Respiratory Medicine, vol. 6, no. 3, p. 223–230, 2018.
- R. Laxminarayan, P. Matsoso, S. Pant, C. Brower, J.-A. Røttingen, K. Klugman and S. Davies, "Access to effective antimicrobials: a worldwide challenge," The Lancet, vol. 387, no. 10014, p. 168–175, 2016.
- L. Say, D. Chou, A. Gemmill, Ö. Tunçalp, A.-B. Moller, J. Daniels, A. M. Gülmezoglu, M. Temmerman and L. Alkema, "Global causes of maternal death: a WHO systematic analysis," The Lancet Global Health, vol. 2, no. 6, pp. e323-e333, 2014.
- 8. R. E. Black, N. Walker, R. Laxminarayan and M. Temmerman, Reproductive, Maternal, Newborn, and Child Health: Disease Control Priorities, Third Edition (Volume 2), R. E. Black, R. Laxminarayan, M. Temmerman and N. Walker, Eds., Washington (DC), 2016.

- National Institute of General Medical Sciences, "Sepsis," 2018. [Online]. Available: https://www. nigms.nih.gov/education/pages/factsheet_sepsis. aspx.
- 10. D. M. Yealy, D. T. Huang, A. Delaney, M. Knight, A. G. Randolph, R. Daniels and T. Nutbeam, "Recognizing and managing sepsis: what needs to be done?," BMC medicine, vol. 13, p. 98, 2015.
- Society of Critical Care Medicine, "Surviving Sepsis Campaign," 2019. [Online]. Available: http://www. survivingsepsis.org/About-SSC/Pages/History.aspx.
- C. Schorr, A. Odden, L. Evans, G. J. Escobar,
 S. Gandhi, S. Townsend and M. Levy, "Implementation of a multicenter performance improvement program for early detection and treatment of severe sepsis in general medical-surgical wards," Journal of hospital medicine, vol. 11 Suppl 1, pp. S32-S39, 2016.
- 13. A. H. Carneiro, P. Póvoa and J. A. Gomes, "Dear Sepsis-3, we are sorry to say that we don't like you," Revista Brasileira de terapia intensiva, vol. 29, no. 1, p. 4–8, 2017.
- 14. M. M. Levy, A. Rhodes, G. S. Phillips, S. R. Townsend, C. A. Schorr, R. Beale, T. Osborn, S. Lemeshow, J.-D. Chiche, A. Artigas and R. P. Dellinger, "Surviving Sepsis Campaign: association between performance metrics and outcomes in a 7.5-year study," Intensive care medicine, vol. 40, no. 11, p. 1623–1633, 2014.
- 15. M. Singer, C. S. Deutschman, C. W. Seymour, M. Shankar-Hari, D. Annane, M. Bauer, R. Bellomo, G. R. Bernard, J.-D. Chiche, C. M. Coopersmith, R. S. Hotchkiss, M. M. Levy, J. C. Marshall, G. S. Martin, S. M. Opal, G. D. Rubenfeld, T. van der Poll, J.-L. Vincent and D. C. Angus, "The Third International Consensus Definitions for Sepsis and Septic Shock (Sepsis-3)," JAMA, vol. 315, no. 8, p. 801–810, 2016.
- 16. A.-M. Georgescu, J. Szederjesi, S.-M. Copotoiu and L. Azamfirei, "Predicting scores correlations in patients with septic shock – a cohort study," Romanian Journal of Anaesthesia and Intensive Care, vol. 21, no. 2, p. 95–98, 2014.

- 17. K.-M. Kaukonen, M. Bailey and R. Bellomo, "Systemic Inflammatory Response Syndrome Criteria for Severe Sepsis," The New England journal of medicine, vol. 373, no. 9, p. 881, 2015.
- C. Lelubre, S. Anselin, K. Zouaoui Boudjeltia,
 P. Biston and M. Piagnerelli, "Interpretation of C-reactive protein concentrations in critically ill patients," BioMed research international, vol. 2013, p. 124021, 2013.
- M. Assicot, C. Bohuon, D. Gendrel, J. Raymond, H. Carsin and J. Guilbaud, "High serum procalcitonin concentrations in patients with sepsis and infection," The Lancet, vol. 341, no. 8844, p. 515–518. 1993.
- 20. Y. Wirz, M. A. Meier, L. Bouadma, C. E. Luyt, M. Wolff, J. Chastre, F. Tubach, S. Schroeder, V. Nobre, D. Annane, K. Reinhart, P. Damas, M. Nijsten, A. Shajiei, D. W. deLange, R. O. Deliberato, C. F. Oliveira, Y. Shehabi, J. A. H. van Oers, A. Beishuizen, A. R. J. Girbes, E. Jong, B. Mueller and P. Schuetz, "Effect of procalcitonin-guided antibiotic treatment on clinical outcomes in intensive care unit patients with infection and sepsis patients: a patient-level meta-analysis of randomized trials," Critical care (London, England), vol. 22, no. 1, p. 191, 2018.
- 21. B. M. Biron, A. Ayala and J. L. Lomas-Neira, "Biomarkers for Sepsis: What Is and What Might Be?," Biomarker insights, vol. 10, no. Suppl 4, p. 7–17, 2015.
- 22. C. Chenevier-Gobeaux, D. Borderie, N. Weiss, T. Mallet-Coste and Y.-E. Claessens, "Presepsin (sCD14-ST), an innate immune response marker in sepsis," Clinica chimica acta; international journal of clinical chemistry, vol. 450, p. 97–103, 2015.
- 23.Y. Okamura and H. Yokoi, "Development of a point-of-care assay system for measurement of presepsin (sCD14-ST)," Clinica chimica acta; international journal of clinical chemistry, vol. 412, no. 23-24, p. 2157–2161, 2011.
- 24. T. Shozushima, G. Takahashi, N. Matsumoto, M. Kojika, Y. Okamura and S. Endo, "Usefulness of presepsin (sCD14-ST) measurements as a marker

- for the diagnosis and severity of sepsis that satisfied diagnostic criteria of systemic inflammatory response syndrome," Journal of infection and chemotherapy: official journal of the Japan Society of Chemotherapy, vol. 17, no. 6, p. 764–769, 2011.
- 25. J. Y. Ham and K. E. Song, "Impact of specimen mixing methods on presepsin point-of-care test results using whole blood," Clinical chemistry and laboratory medicine, vol. 54, no. 5, pp. e151-4, 2016.
- 26. R. Carpio, J. Zapata, E. Spanuth and G. Hess, "Utility of presepsin (sCD14-ST) as a diagnostic and prognostic marker of sepsis in the emergency department," Clinica chimica acta; international journal of clinical chemistry, vol. 450, p. 169–175, 2015.
- 27. E. Spanuth, H. Ebelt, B. Ivandic and K. Werdan, "2.29 Diagnostic and prognostic value of presepsin (soluble CD14 subtype) in emergency patients with early sepsis using the new assay PATHFAST presepsin," in Advances in Clinical Chemistry and Laboratory Medicine, Berlin, Boston, DE GRUYTER, 2012.
- 28. K. Hoshino, Y. Irie, M. Mizunuma, K. Kawano, T. Kitamura and H. Ishikura, "Incidence of elevated procalcitonin and presepsin levels after severe trauma: a pilot cohort study," Anaesthesia and intensive care, vol. 45, no. 5, p. 600–604, 2017.
- 29. S. Masson, P. Caironi, E. Spanuth, R. Thomae, M. Panigada, G. Sangiorgi, R. Fumagalli, T. Mauri, S. Isgrò, C. Fanizza, M. Romero, G. Tognoni, R. Latini and L. Gattinoni, "Presepsin (soluble CD14 subtype) and procalcitonin levels for mortality prediction in sepsis: data from the Albumin Italian Outcome Sepsis trial," Critical care (London, England), vol. 18, no. 1, p. R6, 2014.
- 30. S. Endo, Y. Suzuki, G. Takahashi, T. Shozushima, H. Ishikura, A. Murai, T. Nishida, Y. Irie, M. Miura, H. Iguchi, Y. Fukui, K. Tanaka, T. Nojima and Y. Okamura, "Presepsin as a powerful monitoring tool for the prognosis and treatment of sepsis: a multicenter prospective study," Journal of infection and chemotherapy: official journal of the Japan Society of Chemotherapy, vol. 20, no. 1, p. 30–34, 2014.

- 31. S. Masson, P. Caironi, C. Fanizza, R. Thomae, R. Bernasconi, A. Noto, R. Oggioni, G. S. Pasetti, M. Romero, G. Tognoni, R. Latini and L. Gattinoni, "Erratum to: Circulating presepsin (soluble CD14 subtype) as a marker of host response in patients with severe sepsis or septic shock: data from the multicenter, randomized ALBIOS trial," Intensive care medicine, vol. 41, no. 9, p. 1736, 2015.
- 32. T. Nagata, Y. Yasuda, M. Ando, T. Abe, T. Katsuno, S. Kato, N. Tsuboi, S. Matsuo and S. Maruyama, "Clinical impact of kidney function on presepsin levels," PloS one, vol. 10, no. 6, p. e0129159, 2015.
- 33. J. Shiota, N. Ohura, S. Higashikawa, T. Yamato, H. Kasahara, K. Itatani and H. Tagawa, "Presepsin as a predictor of critical colonization in CLI hemodialysis patients," Wound repair and regeneration: official publication of the Wound Healing Society [and] the European Tissue Repair Society, vol. 24, no. 1, p. 189–194, 2016.
- 34. G. Takahashi, S. Shibata, Y. Fukui, Y. Okamura and Y. Inoue, "Diagnostic accuracy of procalcitonin and presepsin for infectious disease in patients with acute kidney injury," Diagnostic microbiology and infectious disease, vol. 86, no. 2, p. 205–210, 2016.
- 35. K. Takeda and S. Akira, "Roles of Toll-like receptors in innate immune responses," Genes to cells: devoted to molecular & cellular mechanisms, vol. 6, no. 9, p. 733–742, 2001.
- 36. R. Dziarski and D. Gupta, "Function of CD14 as a peptidoglycan receptor: differences and similarities with LPS," Journal of Endotoxin Research, vol. 5, no. 1-2, p. 56–61, 1999.
- D. Giavarina and M. Carta, "Determination of reference interval for presepsin, an early marker for sepsis," Biochemia medica, vol. 25, no. 1, p. 64–68, 2015.
- 38. S. Endo, Y. Suzuki, G. Takahashi, T. Shozushima, H. Ishikura, A. Murai, T. Nishida, Y. Irie, M. Miura, H. Iguchi, Y. Fukui, K. Tanaka, T. Nojima and Y. Okamura, "Usefulness of presepsin in the diagnosis of sepsis in a multicenter prospective study," Journal of infection and chemotherapy: official journal of the Japan Society of Chemotherapy, vol. 18, no. 6, p. 891–897, 2012.

- 39. B. Lu, Y. Zhang, C. Li, C. Liu, Y. Yao, M. Su and S. Shou, "The utility of presepsin in diagnosis and risk stratification for the emergency patients with sepsis," The American journal of emergency medicine, vol. 36, no. 8, p. 1341–1345, 2018.
- 40.T. Yamamoto, T. Nishimura, S. Kaga, K. Uchida, Y. Tachibana, M. Esaki, W. Fukushima, K. Kondo and Y. Mizobata, "Diagnostic accuracy of presepsin for sepsis by the new Sepsis-3 definitions," The American journal of emergency medicine, 2019.
- 41. X. Tong, Y. Cao, M. Yu and C. Han, "Presepsin as a diagnostic marker for sepsis: evidence from a bivariate meta-analysis," Therapeutics and clinical risk management, vol. 11, p. 1027–1033, 2015.
- 42. J. Wu, L. Hu, G. Zhang, F. Wu and T. He, "Accuracy of Presepsin in Sepsis Diagnosis: A Systematic Review and Meta-Analysis," PloS one, vol. 10, no. 7, p. e0133057, 2015.
- 43. X. Zhang, D. Liu, Y.-N. Liu, R. Wang and L.-X. Xie, "The accuracy of presepsin (sCD14-ST) for the diagnosis of sepsis in adults: a meta-analysis," Critical care (London, England), vol. 19, p. 323, 2015.
- 44. Z. Zheng, L. Jiang, L. Ye, Y. Gao, L. Tang and M. Zhang, "The accuracy of presepsin for the diagnosis of sepsis from SIRS: a systematic review and meta-analysis," Annals of intensive care, vol. 5, no. 1, p. 48, 2015.
- 45. J. Zhang, Z.-D. Hu, J. Song and J. Shao, "Diagnostic Value of Presepsin for Sepsis: A Systematic Review and Meta-Analysis," Medicine, vol. 94, no. 47, p. e2158, 2015.
- 46. Y. Liu, J.-H. Hou, Q. Li, K.-J. Chen, S.-N. Wang and J.-M. Wang, "Biomarkers for diagnosis of sepsis in patients with systemic inflammatory response syndrome: a systematic review and meta-analysis," SpringerPlus, vol. 5, no. 1, p. 2091, 2016.
- 47. C.-C. Wu, H.-M. Lan, S.-T. Han, C.-H. Chaou, C.-F. Yeh, S.-H. Liu, C.-H. Li, G. N. Blaney, Z.-Y. Liu and K.-F. Chen, "Comparison of diagnostic accuracy in sepsis between presepsin, procalcitonin, and Greactive protein: a systematic review and meta-analysis," Annals of intensive care, vol. 7, no. 1, p. 91, 2017. J Infect Chemother. 17 (2011):764-9

- 48. Y. Kondo, Y. Umemura, K. Hayashida, Y. Hara, M. Aihara and K. Yamakawa, "Diagnostic value of procalcitonin and presepsin for sepsis in critically ill adult patients: a systematic review and meta-analysis," Journal of intensive care, vol. 7, p. 22, 2019.
- 49. J. Rabensteiner, M. Skvarc, M. Hoenigl, J. Osredkar, F. Prueller, M. Reichsoellner, R. Krause and R. B. Raggam, "Diagnostic and prognostic potential of presepsin in Emergency Department patients presenting with systemic inflammatory response syndrome," The Journal of infection, vol. 69, no. 6, p. 627–630, 2014.
- 50. M. Ugajin, Y. Matsuura, K. Matsuura and H. Matsuura, "Impact of initial plasma presepsin level for clinical outcome in hospitalized patients with pneumonia," Journal of thoracic disease, vol. 11, no. 4, p. 1387–1396, 2019.
- 51. Y. Bamba, H. Moro, N. Aoki, T. Koizumi, Y. Ohshima, S. Watanabe, T. Sakagami, T. Koya, T. Takada and T. Kikuchi, "Increased presepsin levels are associated with the severity of fungal bloodstream infections," PloS one, vol. 13, no. 10, p. e0206089, 2018.
- 52. Z.-J. Qi, H. Yu, J. Zhang and C.-S. Li, "Presepsin as a novel diagnostic biomarker for differentiating active pulmonary tuberculosis from bacterial community acquired pneumonia," Clinica chimica acta; international journal of clinical chemistry, vol. 478, p. 152–156, 2018.
- 53. Y. Arai, K. Mizugishi, K. Nonomura, K. Naitoh, A. Takaori-Kondo and K. Yamashita, "Phagocytosis by human monocytes is required for the secretion of presepsin," Journal of infection and chemotherapy: official journal of the Japan Society of Chemotherapy, vol. 21, no. 8, p. 564–569, 2015.
- 54. H. Yamaguchi, A. Takuma, E. Fukuoka, H. Oto, M. Inoue, T. Nagahama, S. Fukuoka, T. Soga, Y. Umeda and S. Kimura, "Concentration of Soluble CD14-Subtype (Presepsin) in Plasma of Non-Inflammatory and Septic Febrile Children," Rinsho byori. The Japanese journal of clinical pathology, vol. 64, no. 9, p. 1001–1006, 2016.

- 55. G. Lippi and G. Cervellin, "Can presepsin be used for screening invasive fungal infections?," Annals of translational medicine, vol. 7, no. 5, p. 87, 2019.
- 56. Y. Koizumi, K. Shimizu, M. Shigeta, T. Okuno, H. Minamiguchi, K. Kito, K. Hodohara, Y. Yamagishi, A. Andoh, Y. Fujiyama and H. Mikamo, "Plasma presepsin level is an early diagnostic marker of severe febrile neutropenia in hematologic malignancy patients," BMC infectious diseases, vol. 17, no. 1, p. 27, 2017.
- 57. K. Ebisawa, J. Koya, K. Nakazaki, S. Arai, F. Nakamura and M. Kurokawa, "Usefulness of presepsin for early detection of infections in patients with hematologic disorders," Clinica chimica acta; international journal of clinical chemistry, vol. 486, p. 374–380, 2018.
- 58. S. Nanno, H. Koh, T. Katayama, M. Hashiba, A. Sato, Y. Makuuchi, J. Nagasaki, M. Kuno, T. Yoshimura, H. Okamura, M. Nishimoto, A. Hirose, M. Nakamae, T. Nakane, M. Hino and H. Nakamae, "Plasma Levels of Presepsin (Soluble CD14-subtype) as a Novel Prognostic Marker for Hemophagocytic Syndrome in Hematological Malignancies," Internal medicine (Tokyo, Japan), vol. 55, no. 16, p. 2173–2184, 2016.
- 59. D. Popov, M. Plyushch, S. Ovseenko, M. Abramyan, O. Podshchekoldina and M. Yaroustovsky, "Prognostic value of sCD14-ST (presepsin) in cardiac surgery," Kardiochirurgia i torakochirurgia polska = Polish journal of cardio-thoracic surgery, vol. 12, no. 1, p. 30–36, 2015.
- 60. H. Bomberg, M. Klingele, S. Wagenpfeil, E. Spanuth, T. Volk, D. I. Sessler, H.-J. Schäfers and H. V. Groesdonk, "Presepsin (sCD14-ST) Is a Novel Marker for Risk Stratification in Cardiac Surgery Patients," Anesthesiology, vol. 126, no. 4, p. 631–642, 2017.5
- 61. G. Novelli, V. Morabito, G. Ferretti, F. Pugliese, F. Ruberto, F. Venuta, L. Poli, M. Rossi and P. B. Berloco, "Pathfast presepsin assay for early diagnosis of bacterial infections in surgical patients: preliminary study," Transplantation proceedings, vol. 45, no. 7, p. 2750–2753, 2013.

- 62. K. Klouche, J. P. Cristol, J. Devin, V. Gilles, N. Kuster, R. Larcher, L. Amigues, P. Corne, O. Jonquet and A. M. Dupuy, "Diagnostic and prognostic value of soluble CD14 subtype (Presepsin) for sepsis and community-acquired pneumonia in ICU patients," Annals of intensive care, vol. 6, no. 1, p. 59, 2016.
- 63. J. Lin, H. Sun, J. Li, Y. Zheng, C. Shao, Y. H. Zhang and H. Chang, "Role of Presepsin for the Assessment of Acute Cholangitis Severity," Clinical laboratory, vol. 62, no. 4, p. 679–687, 2016.
- 64. T. Imagama, A. Tokushige, K. Seki, T. Seki, D. Nakashima, H. Ogasa, T. Sakai and T. Taguchi, "Early diagnosis of septic arthritis using synovial fluid presepsin: A preliminary study," Journal of infection and chemotherapy: official journal of the Japan Society of Chemotherapy, vol. 25, no. 3, p. 170–174, 2019.
- 65. N. Watanabe, T. Ishii, N. Kita, N. Kanaji, H. Nakamura, N. Nanki, Y. Ueda, Y. Tojo, N. Kadowaki and S. Bandoh, "The usefulness of pleural fluid presepsin, C-reactive protein, and procalcitonin in distinguishing different causes of pleural effusions," BMC pulmonary medicine, vol. 18, no. 1, p. 176, 2018.
- 66. S. Tanimura, Y. Fujieda, M. Kono, Y. Shibata, R. Hisada, E. Sugawara, H. Nakamura, K. Ohmura, S. Shimamura, A. Mitani, H. Shida, T. Watanabe, M. Kato, K. Oku, T. Bohgaki, O. Amengual, S. Yasuda, C. Shimizu and T. Atsumi, "Clinical significance of plasma presepsin levels in patients with systemic lupus erythematosus," Modern rheumatology, vol. 28, no. 5, p. 865–871, 2018.
- 67. J. Lin, Z. Li, Y. Zheng, Y. Zhang, C. Shao, G. Liu and J. Li, "Elevated Presepsin Levels are Associated with Severity and Prognosis of Severe Acute Pancreatitis," Clinical laboratory, vol. 62, no. 9, p. 1699–1708, 2016.
- 68. F. N. T. Caglar, N. Isiksacan, I. Biyik, S. Opan, H. Cebe and I. F. Akturk, "Presepsin (sCD14-ST): could it be a novel marker for the diagnosis of ST elevation myocardial infarction?," Archives of medical sciences. Atherosclerotic diseases, vol. 2, no. 1, pp. e3-e8, 2017.

- 69. L. Pugni, C. Pietrasanta, S. Milani, C. Vener, A. Ronchi, M. Falbo, M. Arghittu and F. Mosca, "Presepsin (Soluble CD14 Subtype): Reference Ranges of a New Sepsis Marker in Term and Preterm Neonates," PloS one, vol. 10, no. 12, p. e0146020, 2015.
- C. Poggi, T. Bianconi, E. Gozzini, M. Generoso and C. Dani, "Presepsin for the detection of late-onset sepsis in preterm newborns," Pediatrics, vol. 135, no. 1, p. 68–75, 2015.
- 71. P. Montaldo, R. Rosso, A. Santantonio, G. Chello and P. Giliberti, "Presepsin for the detection of early-onset sepsis in preterm newborns," Pediatric research, vol. 81, no. 2, p. 329–334, 2017.
- 72. I. Bellos, G. Fitrou, V. Pergialiotis, N. Thomakos, D. N. Perrea and G. Daskalakis, "The diagnostic accuracy of presepsin in neonatal sepsis: a metaanalysis," European journal of pediatrics, vol. 177, no. 5, p. 625–632, 2018.
- 73. B. Jovanovic, O. Djuric, L. Markovic-Denic, A. Isakovic, K. Doklestic, S. Stankovic, S. Vidicevic, I. Palibrk, J. Samardzic and V. Bumbasirevic, "Prognostic value of presepsin (soluble CD14-subtype) in diagnosis of ventilator-associated pneumonia and sepsis in trauma patients," Vojnosanitetski pregled, vol. 75, no. 10, p. 968–977, 2018.
- 74. X. Song, Y. Song, Y. Yuan, P. Zhang and X. Zhang, "Prognostic value of presepsin for outcomes and complications in enterocutaneous fistula complicated by abdominal sepsis," International journal of surgery (London, England), vol. 33 Pt A, p. 96–101, 2016.
- 75. L. Ruan, G.-Y. Chen, Z. Liu, Y. Zhao, G.-Y. Xu, S.-F. Li, C.-N. Li, L.-S. Chen and Z. Tao, "The combination of procalcitonin and C-reactive protein or presepsin alone improves the accuracy of diagnosis of neonatal sepsis: a meta-analysis and systematic review," Critical care (London, England), vol. 22, no. 1, p. 316, 2018.
- 76. F. T. Ali, M. A. M. Ali, M. M. Elnakeeb and H. N. M. Bendary, "Presepsin is an early monitoring biomarker for predicting clinical outcome in patients with sepsis," Clinica chimica acta; international journal of clinical chemistry, vol. 460, p. 93–101, 2016.

- 77. C. Balcl, H. Sungurtekin, E. Gürses, U. Sungurtekin and B. Kaptanoglu, "Usefulness of procalcitonin for diagnosis of sepsis in the intensive care unit," Critical Care, vol. 7, no. 1, p. 85, 2002.
- 78. P. R. Bauer, R. Kashyap, S. C. League, J. G. Park, D. R. Block, N. A. Baumann, A. Algeciras-Schimnich, S. M. Jenkins, C. Y. Smith, O. Gajic and R. S. Abraham, "Diagnostic accuracy and clinical relevance of an inflammatory biomarker panel for sepsis in adult critically ill patients," Diagnostic microbiology and infectious disease, vol. 84, no. 2, p. 175–180, 2016.
- 79. M. Behnes, T. Bertsch, D. Lepiorz, S. Lang, F. Trinkmann, M. Brueckmann, M. Borggrefe and U. Hoffmann, "Diagnostic and prognostic utility of soluble CD 14 subtype (presepsin) for severe sepsis and septic shock during the first week of intensive care treatment," Critical care (London, England), vol. 18, no. 5, p. 507, 2014.
- 80. Ö. Cakır Madenci, S. Yakupoğlu, N. Benzonana, N. Yücel, D. Akbaba and A. Orçun Kaptanağası, "Evaluation of soluble CD14 subtype (presepsin) in burn sepsis," Burns: journal of the International Society for Burn Injuries, vol. 40, no. 4, p. 664–669, 2014.
- 81. A. Enguix-Armada, R. Escobar-Conesa, A. La García-De Torre and M. V. La Torre-Prados, "Usefulness of several biomarkers in the management of septic patients: C-reactive protein, procalcitonin, presepsin and mid-regional pro-adrenomedullin," Clinical chemistry and laboratory medicine, vol. 54, no. 1, p. 163–168, 2016.
- 82. S. Gibot, M.-N. Kolopp-Sarda, M. C. Béné, A. Cravoisy, B. Levy, G. C. Faure and P.-E. Bollaert, "Plasma level of a triggering receptor expressed on myeloid cells-1: its diagnostic accuracy in patients with suspected sepsis," Annals of internal medicine, vol. 141, no. 1, p. 9–15, 2004.
- 83. M. Godnic, D. Stubljar, D. Stubjar, M. Skvarc and T. Jukic, "Diagnostic and prognostic value of sCD14-ST-presepsin for patients admitted to hospital intensive care unit (ICU)," Wiener klinische Wochenschrift, vol. 127, no. 13-14, p. 521–527, 2015.

- 84. C. Leli, M. Ferranti, U. Marrano, Z. S. Al Dhahab, S. Bozza, E. Cenci and A. Mencacci, "Diagnostic accuracy of presepsin (sCD14-ST) and procalcitonin for prediction of bacteraemia and bacterial DNAaemia in patients with suspected sepsis," Journal of medical microbiology, vol. 65, no. 8, p. 713–719, 2016.
- 85. F. Miglietta, M. L. Faneschi, G. Lobreglio, C. Palumbo, A. Rizzo, M. Cucurachi, G. Portaccio, F. Guerra and M. Pizzolante, "Procalcitonin, G-reactive protein and serum lactate dehydrogenase in the diagnosis of bacterial sepsis, SIRS and systemic candidiasis," Le infezioni in medicina, vol. 23, no. 3, p. 230–237, 2015.
- 86. L. G. d. G. Romualdo, P. E. Torrella, M. V. González, R. J. Sánchez, A. H. Holgado, A. O. Freire, S. R. Acebes and M. D. A. Otón, "Diagnostic accuracy of presepsin (soluble CD14 subtype) for prediction of bacteremia in patients with systemic inflammatory response syndrome in the Emergency Department," Clinical biochemistry, vol. 47, no. 7-8, p. 505–508, 2014.
- 87. O. Selberg, H. Hecker, M. Martin, A. Klos, W. Bautsch and J. Köhl, "Discrimination of sepsis and systemic inflammatory response syndrome by determination of circulating plasma concentrations of procalcitonin, protein complement 3a, and interleukin-6," Critical care medicine, vol. 28, no. 8, p. 2793–2798, 2000.
- 88. W. Takahashi, T.-A. Nakada, M. Yazaki and S. Oda, "Interleukin-6 Levels Act as a Diagnostic Marker for Infection and a Prognostic Marker in Patients with Organ Dysfunction in Intensive Care Units," Shock (Augusta, Ga.), vol. 46, no. 3, p. 254–260, 2016.
- 89. H. Ugarte, E. Silva, D. Mercan, A. Mendonça and J. L. Vincent, "Procalcitonin used as a marker of infection in the intensive care unit," Critical care medicine, vol. 27, no. 3, p. 498–504, 1999.
- 90.P. J. van der Geest, M. Mohseni, J. Linssen, S. Duran, R. Jonge and A. B. J. Groeneveld, "The intensive care infection score a novel marker for the prediction of infection and its severity," Critical care (London, England), vol. 20, no. 1, p. 180, 2016. J Infect Chemother. 2019 Mar;25(3):170-174. doi: 10.1016/j.jiac.2018.10.015. Epub 2018 Nov 23

- 91. H. R. Wong, C. J. Lindsell, P. Lahni, K. W. Hart and S. Gibot, "Interleukin 27 as a sepsis diagnostic biomarker in critically ill adults," Shock (Augusta, Ga.), vol. 40, no. 5, p. 382–386, 2013.
- 92. Y. Yang, J. Xie, F. Guo, F. Longhini, Z. Gao, Y. Huang and H. Qiu, "Combination of C-reactive protein, procalcitonin and sepsis-related organ failure score for the diagnosis of sepsis in critical patients," Annals of intensive care, vol. 6, no. 1, p. 51, 2016.
- 93. M. Nakamura, T. Takeuchi, K. Naito, K. Shirakawa, Y. Hosaka, F. Yamasaki and S. Furusako, "Early elevation of plasma soluble CD14 subtype, a novel biomarker for sepsis, in a rabbit cecal ligation and puncture model," Critical care (London, England), vol. 12, no. Suppl 2, p. P194, 2008.
- 94. S. A. Abudeev, K. V. Kiselev, N. M. Kruglyakov, K. A. Belousova, I. N. Lobanova, O. V. Parinov, Y. D. Udalov, M. A. Zabelin, A. S. Samoilov, E. Cesnulis, T. Killeen and K. A. Popugaev, "Cerebrospinal Fluid Presepsin As a Marker of Nosocomial Infections of the Central Nervous System: A Prospective Observational Study," Frontiers in neurology, vol. 9, p. 58, 2018.
- 95. N. N. Salina, V. P. Nikulina, R. N. Borisov and M. A. Godkov, "PRESEPSIN AS THE EARLY MARKER OF PURULENT SEPTIC COMPLICATIONS IN PATIENTS WITH SEVERE ACUTE PANCREATITIS," Russian Sklifosovsky Journal "Emergency Medical Care", vol. 7, no. 1, p. 30–36, 2018.
- 96. E. A. Titova, A. R. Eirikh, Z. A. Titova, O. G. Zhgut, S. I. Ivanova, T. N. Zateeva, D. V. Petrova and E. M. Reutskaya, "Change in the presepsin level in hemodialysis patients with pneumonia and sepsis," Bulletin of Medical Science, no. 1(13), p. 73–77, 2019.

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